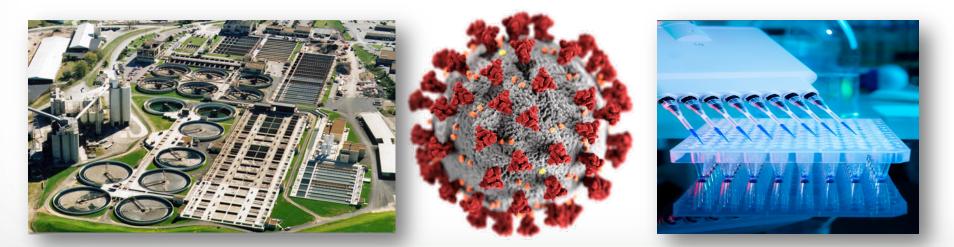


EPA/ORD Method for SARS-CoV-2 Wastewater Monitoring

Nichole Brinkman

US Environmental Protection Agency

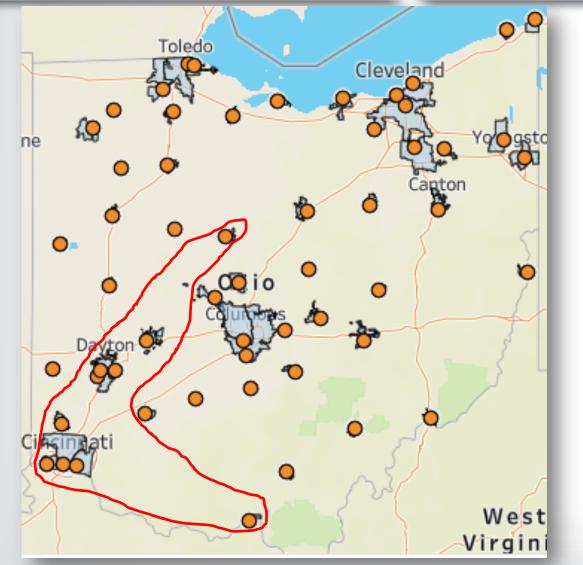
Office of Research and Development



The views expressed in this presentation are those of the author(s) and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency. Any mention of trade names or commercial products does not constitute EPA endorsement or recommendation for use.

1

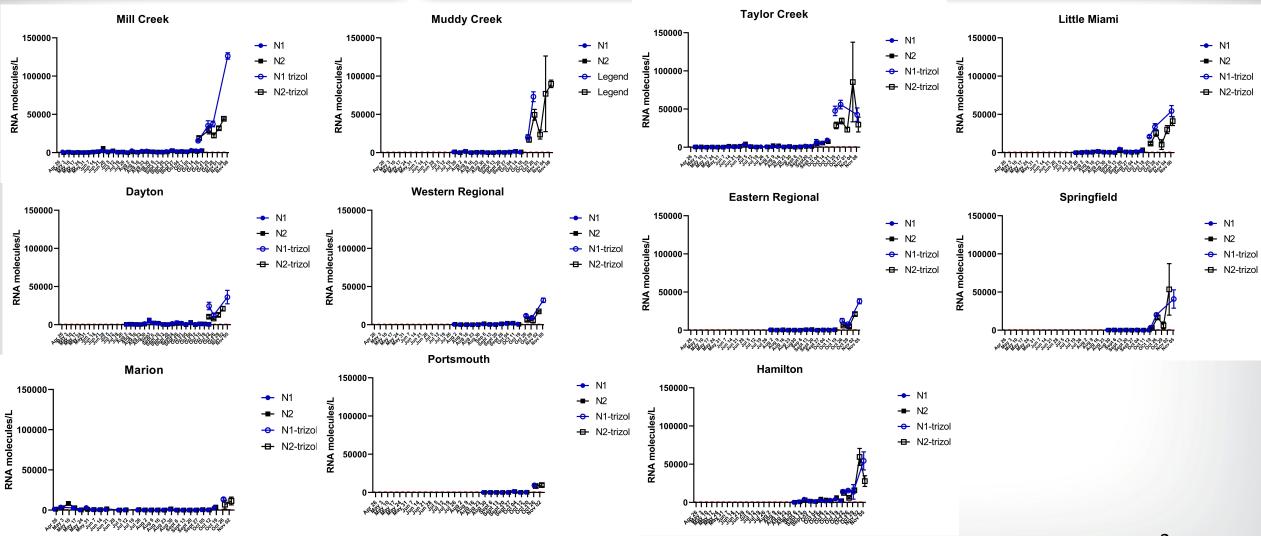
OH Coronavirus Wastewater Monitoring Network



- Initiated in May by Gov. DeWine
- Coordinated by Ohio Water Resources Center at OSU
 - Ohio Department of Health
 - Ohio EPA
 - 56 wastewater utilities
 - University labs
 - Commercial labs
 - USEPA/ORD-Cincinnati

https://coronavirus.ohio.gov/wps/portal/gov/covid-19/dashboards/wastewater

Temporal Trends of SARS-CoV-2 in Sewersheds



SEPA

€PA

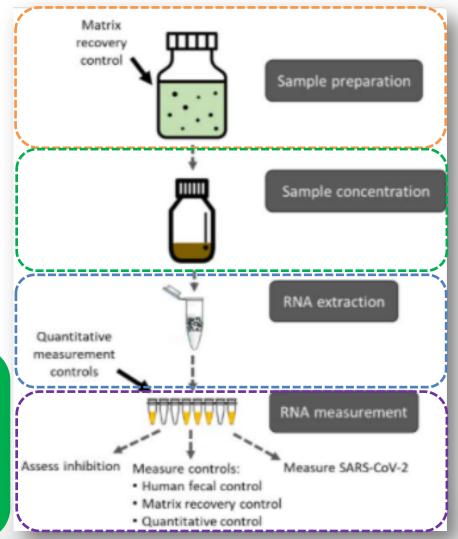
Method Considerations

<u>Sample Type</u> Untreated wastewater Primary sludge Volume

Sample Preparation Storage temperature Homogenization Additives Matrix Spike Clarification

Sample Concentration Ultrafiltration Electronegative membrane filtration Polyethylene glycol (PEG) precipitation

https://medrxiv.org/cgi/content/short/202 0.11.02.20221622v1

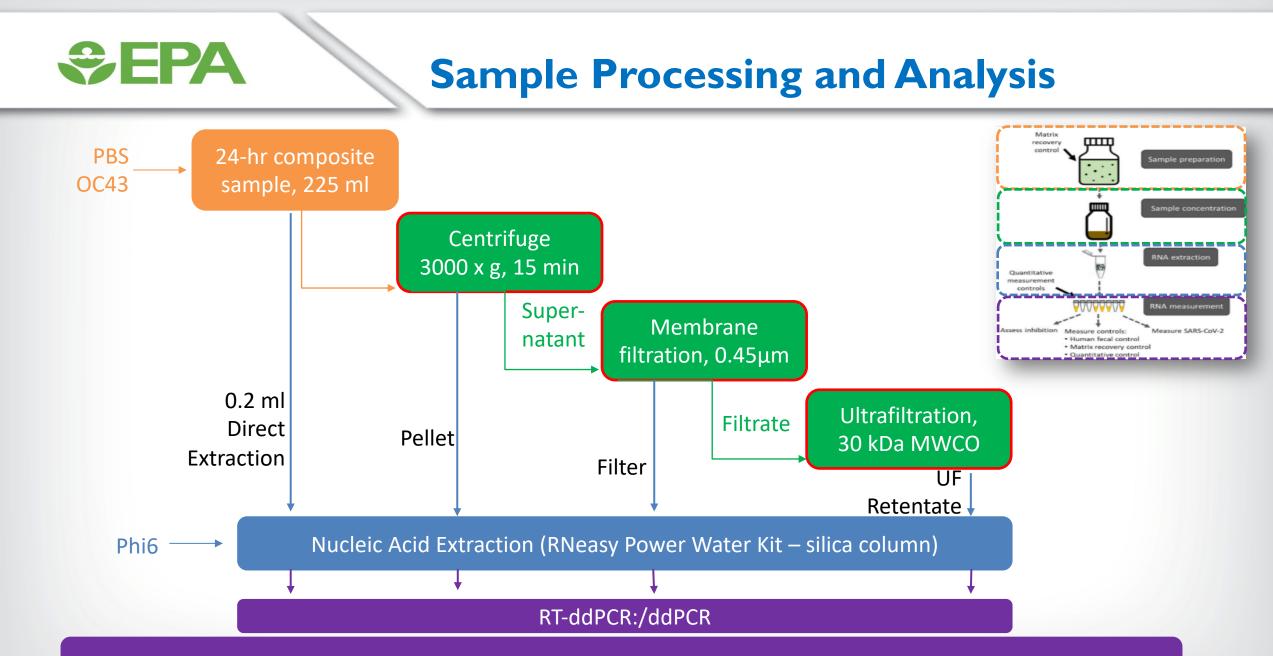


https://www.cdc.gov/coronavirus/2019-ncov/cases-updates/wastewatersurveillance/testing-methods.html Nucleic Acid Extraction Silica columns Magnetic beads Precipitation

RNA/DNA Measurement RT-qPCR RT-ddPCR Genetic targets

Other Considerations Biosafety Supply Chain issues Practicality (time, equipment) QA/QC

https://cwn-rce.ca/covid-19-wastewatercoalition/phase-1-inter-laboratory-study/

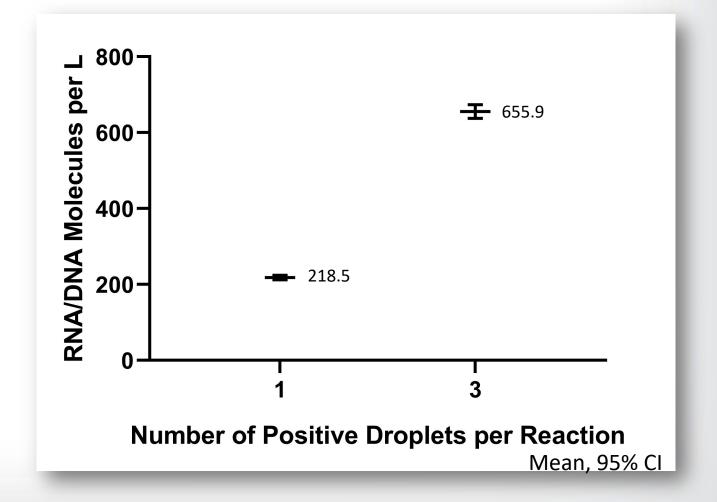


SARS-CoV-2 (N1, N2), RT-ddPCR QC, Inhibition control, Extraction Control, Matrix Recovery Control, Human fecal markers

Limits of Detection/Quantification

- Volume of sample processed
- Concentration factor

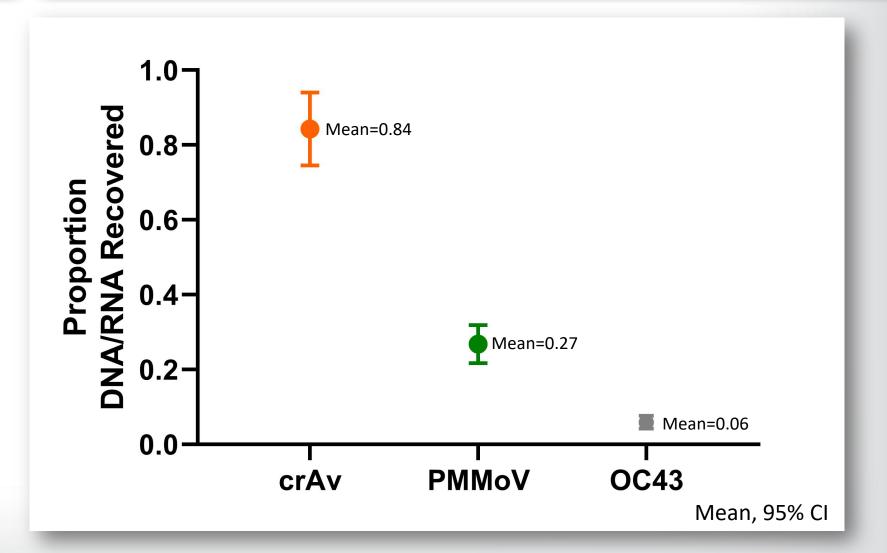
- Volume of processed sample analyzed
- Analytical sensitivity (i.e., minimum detectable concentration)
- Ideal conditions
- Practical limits likely higher due to losses during processing



Recovery Efficiency of Endogenous and Spiked Virus

Endogenous virus

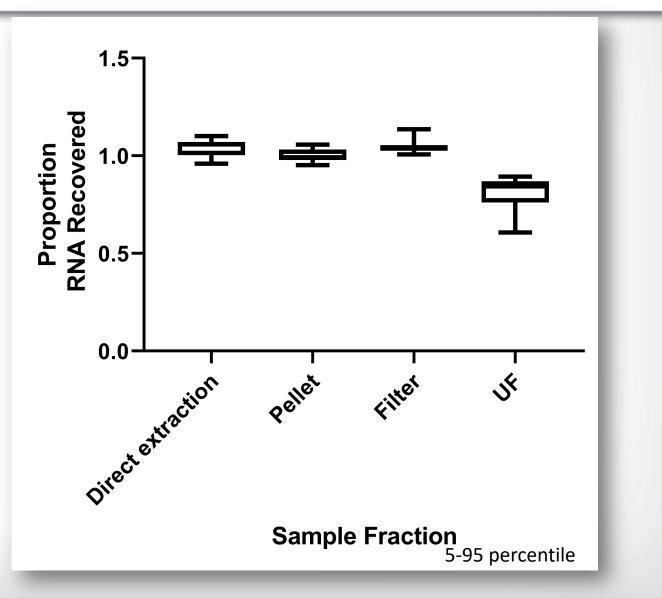
- crAssphage
- Pepper Mild Mottle Virus
- Spiked virus
 - OC43
- Measure concentrations before and after sample processing

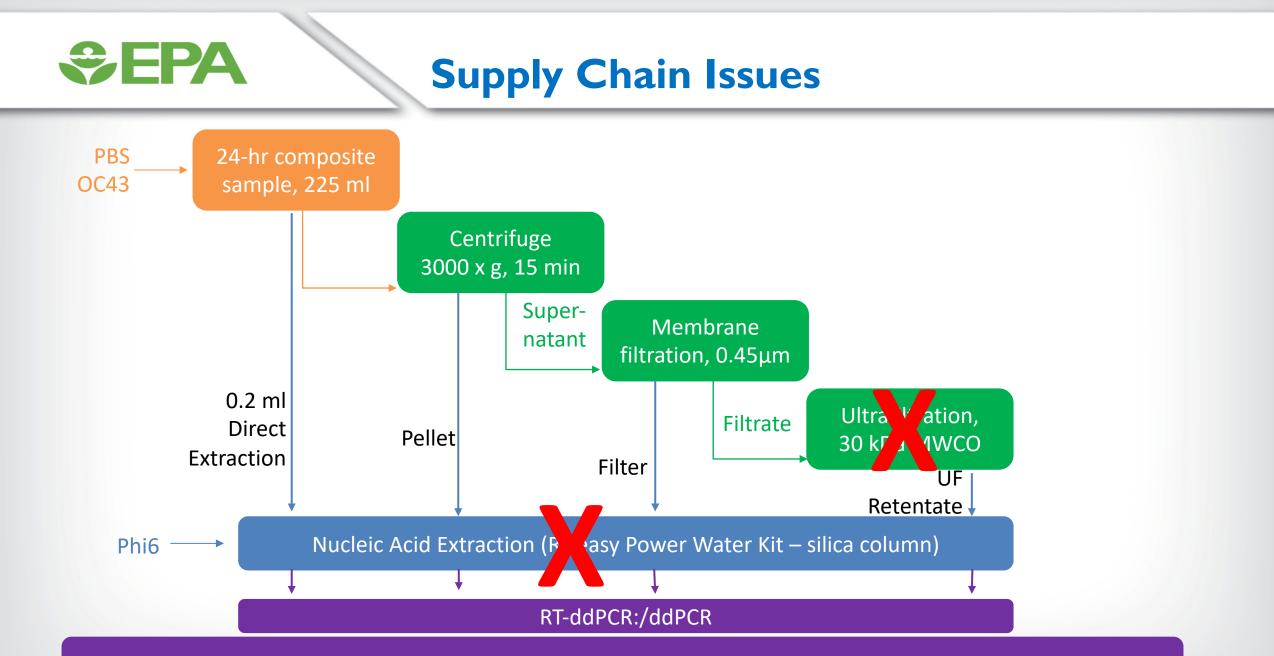


RT-ddPCR Inhibition

• Add RNA before RT-ddPCR

- Compare RNA concentration in sewage sample extracts and matrix-free controls
- Minimal RT-ddPCR inhibition observed



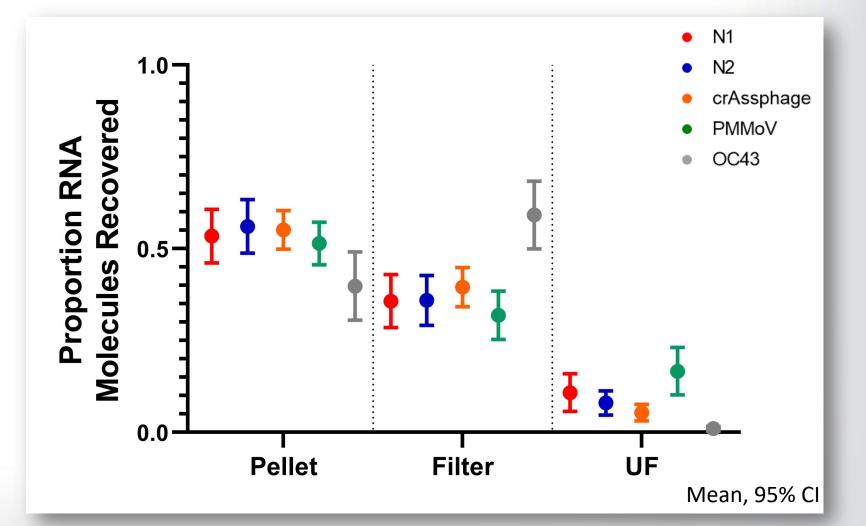


SARS-CoV-2 (N1, N2), RT-ddPCR QC, Inhibition control, Extraction Control, Matrix Recovery Control, Human fecal marker

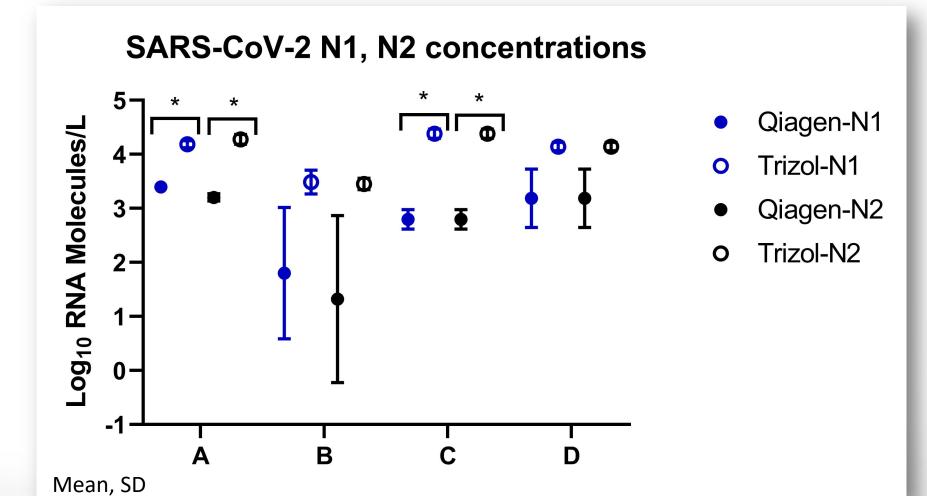
Partitioning of Virus in Sample Fractions

• Where are viruses recovered within samples?

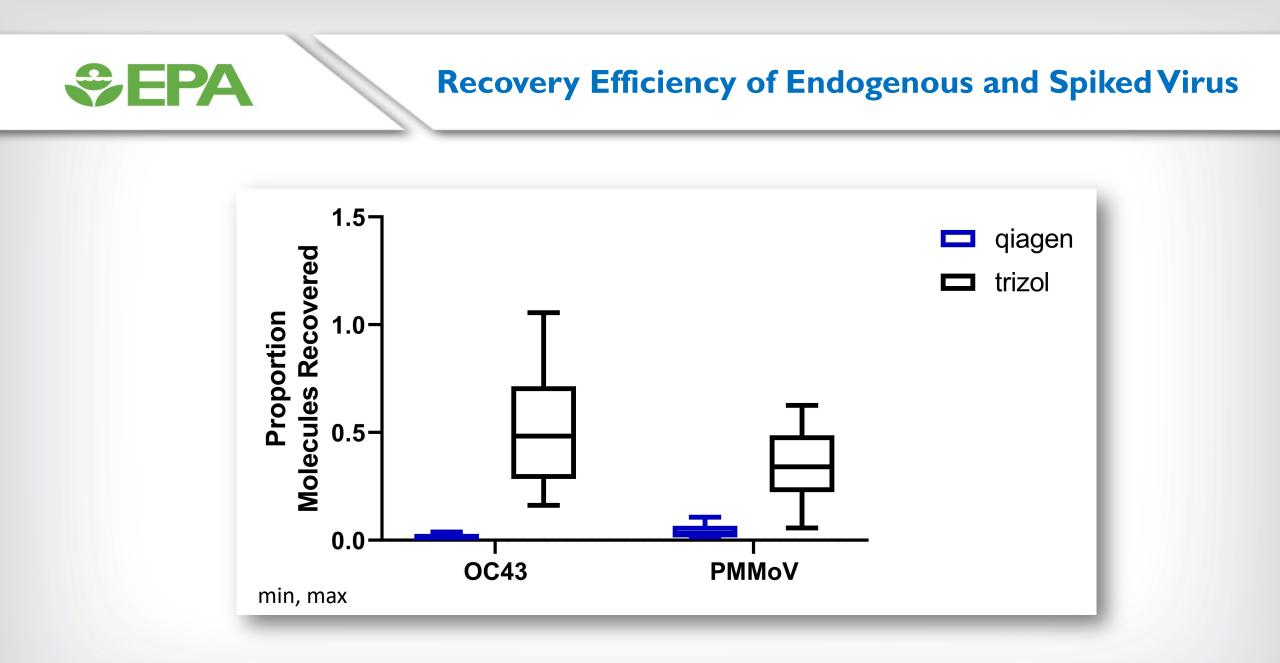
- Proportion of total virus measured in each sample fraction
- ~ 90% measurable virus in pellet and filter fractions



Trizol-chloroform and RNA precipitation

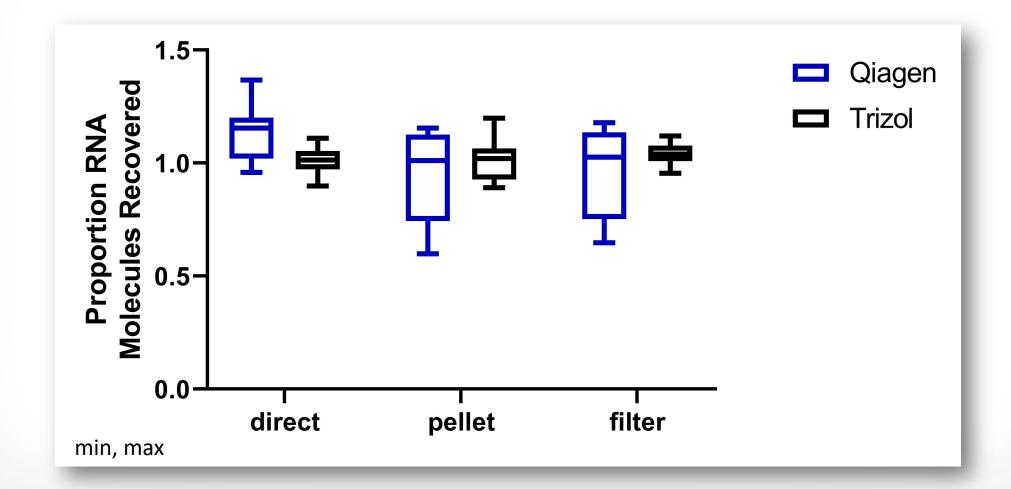


SEPA



€PA

RT-ddPCR Inhibition

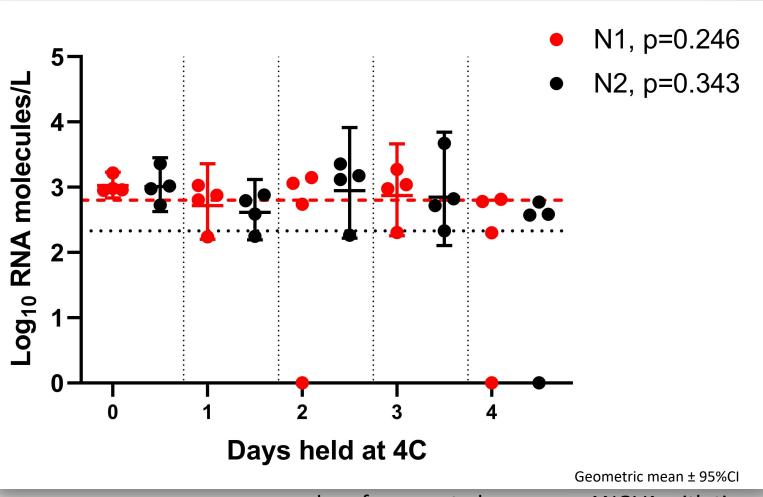


Sample Storage at 4°C

- **24-** hour composite
- Shipped overnight

SEPA

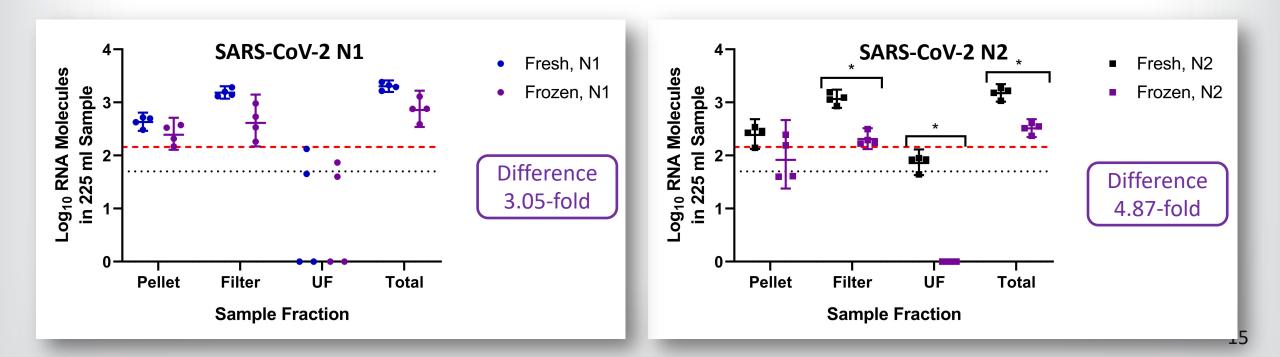
- How long can samples be stored before a significant decrease in viral RNA is observed?
- No significant difference in SARS-CoV-2 RNA up to 4 days at 4°C



p-values for repeated measures ANOVA with time

Sample Storage at -70°C

- What is the effect of freeze-thaw cycle on detection of SARS-CoV-2 RNA?
- Sample collected and processed immediately
- Subsample frozen at -70°C, thawed at 37°C







- Wastewater is a complex and variable matrix
- No standard method for measuring SARS-CoV-2, but many options available
- Quality Control important for assessing method performance and matrix variability
- Supply chain can be disrupted
- Sample handling conditions need more data
 - Temporary storage no reduction at 4°C up to 4 days
 - Long-term storage up to 5-fold reduction after freezing at -80 °C
- Continue to measure SARS-CoV-2 in wastewater at specified sites
- Focus on trends or significant changes in the concentration of viral RNA detected
- Continue to work with ODH to interpret trends for appropriate public health applications

*₽***EPA**

Research Team and Partners

EPA/ORD

Emily Wheaton Maitreyi Nagarkar Chloe Hart Scott Keely Michael Jahne **Alison Franklin** Eunice Varughese Jay Garland Laura Boczek **Randy Revetta Brian Morris** Dave Feldhake Ana Braam **Barry Wiechman** Sarah Okrum Jacob Botkins Leah Julifs

Utilities Metropolitan Sewer District of Greater Cincinnati **Bruce Smith City of Dayton** Chris Clark. Walter Schroder **City of Marion Steve Morris City of Portsmouth Tommy Stewart Montgomery County** lim Davis **City of Hamilton** Mark Smith **City of Springfield** Jeff Yinger

Hamilton County Public Health Department

Chris Griffith

Ohio Water Resources Center

Zuzana Bohrerova

Ohio Department of Health

Rebecca Fugitt

<u>Ohio EPA</u>

Brian Hall

Tiffani Kavalec

University Labs

Ohio State University University of Toledo Kent State University University of Akron